

# Existing Tools For Customized Model Generation Solutions

## Pros & Cons About The Different Methodologies



**Dr. Benedikt Wefers**  
Scientific Program Manager - Custom Model Generation Solutions

### Key responsibilities:

Develop and articulate technical and business solutions for custom model generation by partnering with our customers to understand both project and program goals

# Custom Model Generation

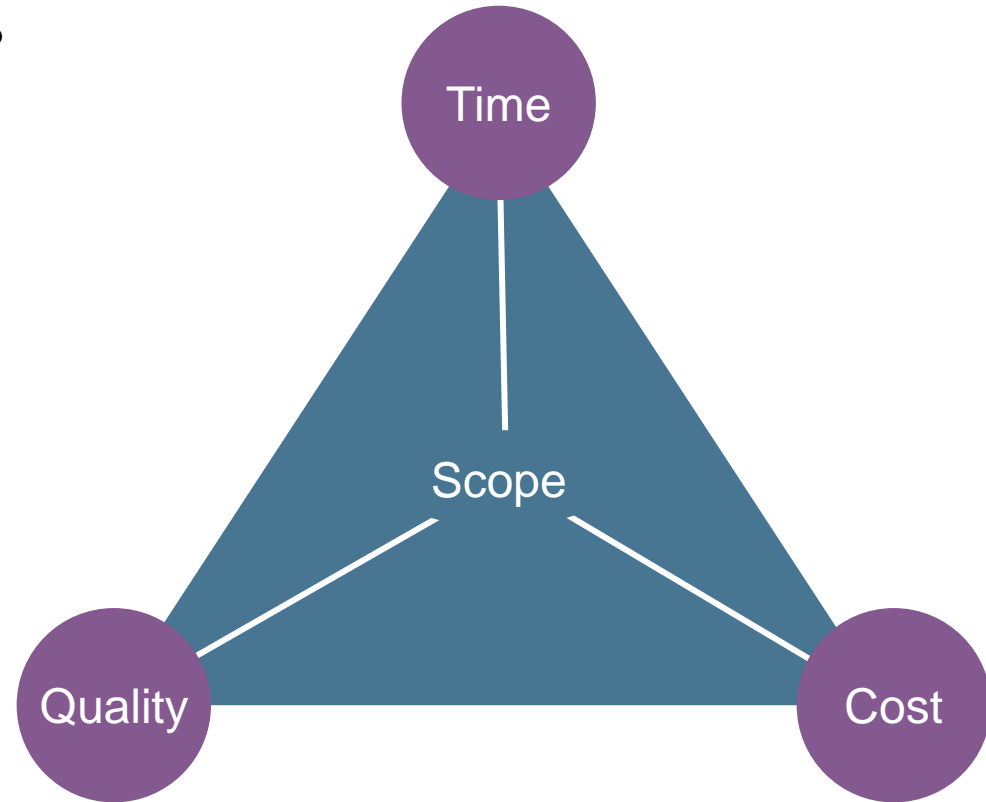
Conceptualize, Strategize and Design

Conceptualize: **Why** do you need the mouse model?

- ▶ Start with the **research goal**
- ▶ Consider downstream analyses

Strategize: **What** is the most appropriate strategy?

- ▶ Evaluate **biological risks**
- ▶ Consider **constraints**



# Custom Model Generation

Conceptualize, Strategize and Design

Conceptualize: **Why** do you need the mouse model?

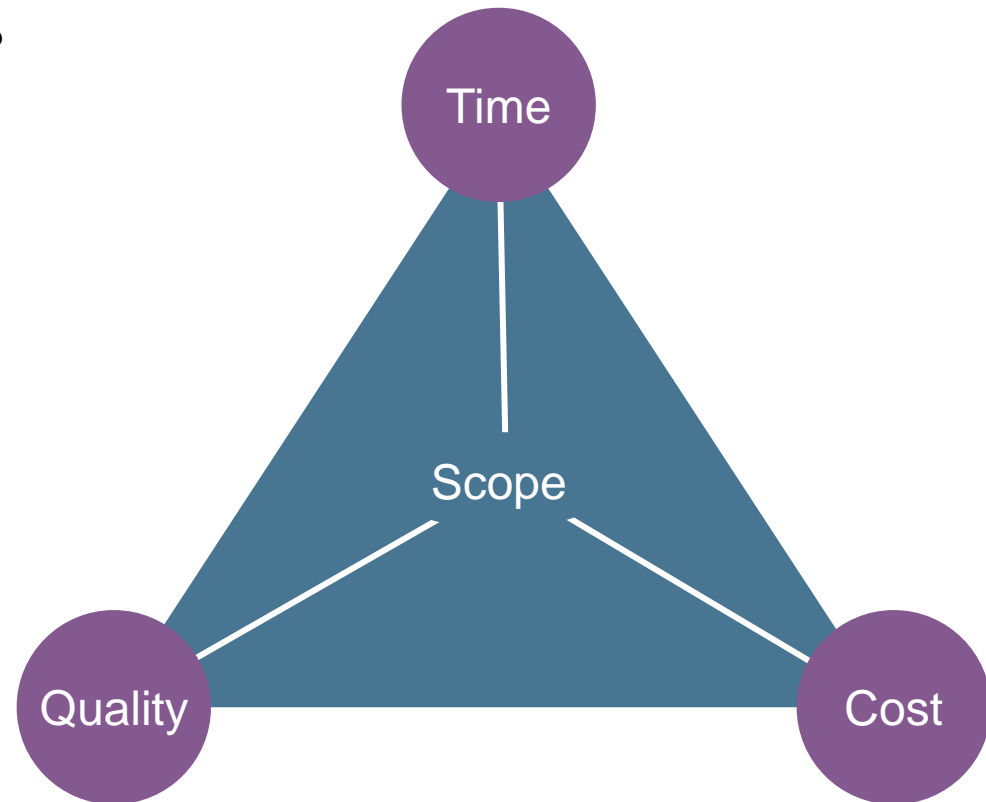
- ▶ Start with the **research goal**
- ▶ Consider downstream analyses

Strategize: **What** is the most appropriate strategy?

- ▶ Evaluate **biological risks**
- ▶ Consider **constraints**

Design: **How** do you make the model?

- ▶ Evaluate **feasibility and efficiency**
- ▶ Consider **technical risks**
- ▶ Consider flexibility
- ▶ Consider tolerances to budget
- ▶ Consider tolerances to timeline

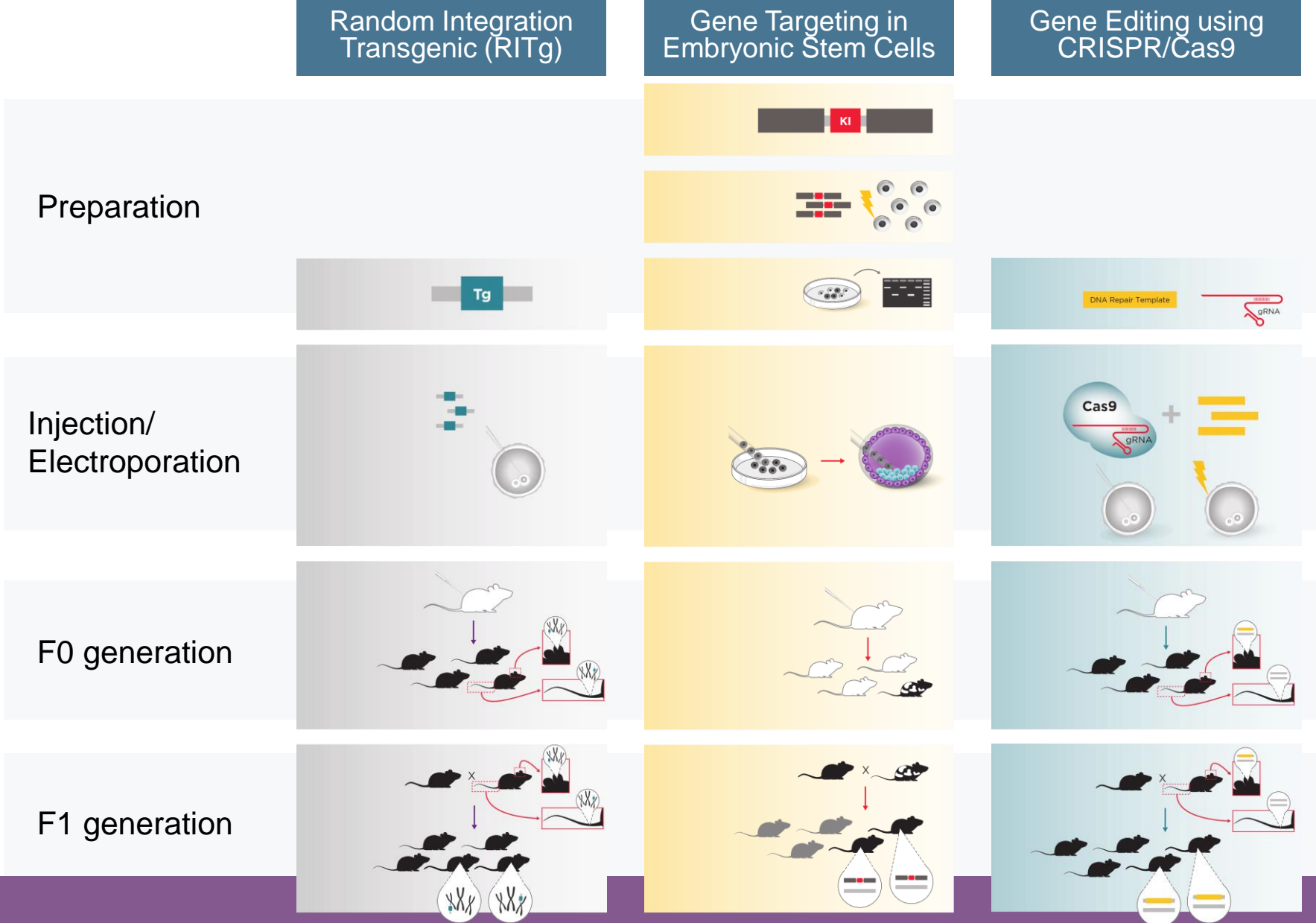


# Outline

- ▶ Major Methods for Genome Modification
  - ▶ Random Integration Transgenics (RITg)
  - ▶ Gene Targeting in Embryonic Stem Cells (ESC)
  - ▶ Gene Editing in Zygotes using CRISPR/Cas9
- ▶ Advancing Model Generation



# Three Major Methods for Modifying the Genome



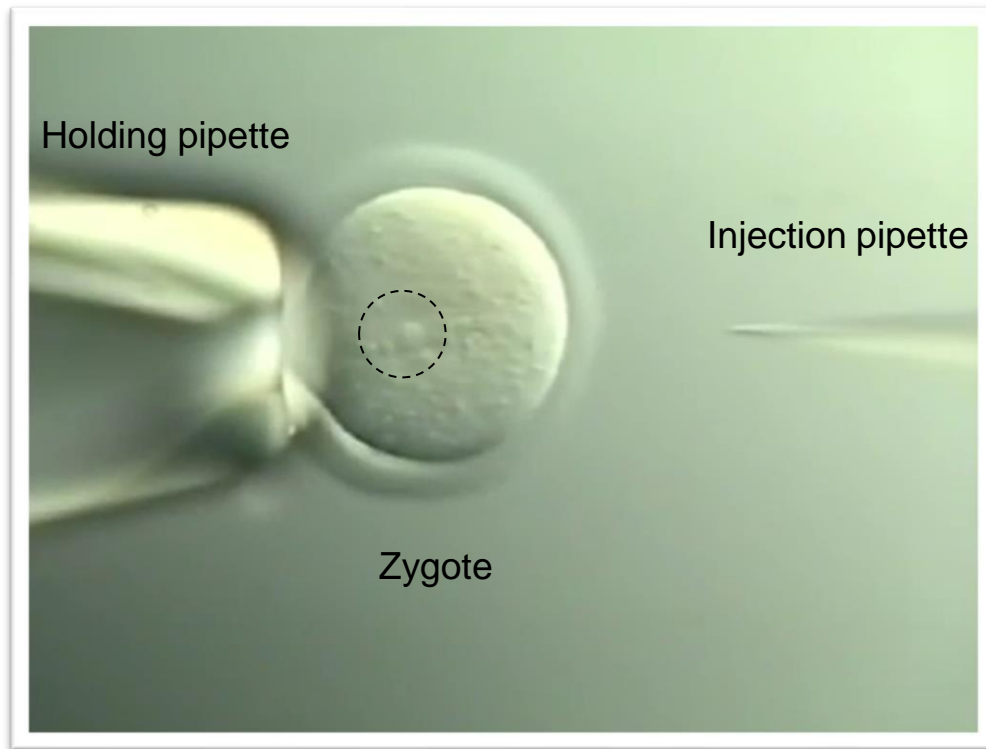
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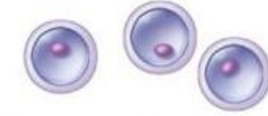


# Random Integration Transgenic Mice (RITg)

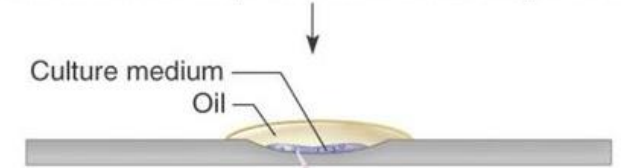
## Model Generation Process



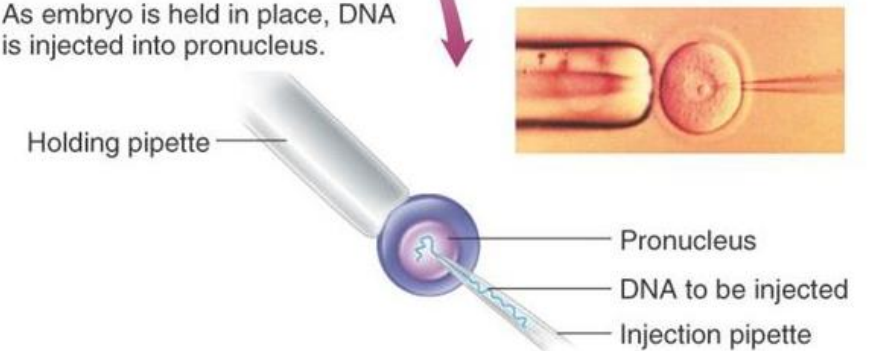
Several embryos recovered from female



Embryos transferred to a depression slide containing culture medium



As embryo is held in place, DNA is injected into pronucleus.



Several injected embryos are placed into oviduct of receptive female.



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# Random Integration Transgenic Mice (RITg)

## Model Generation Process



### Major Advantages

- ▶ Wide range of transgene designs possible (e.g., promoter choice, tags or reporters)
- ▶ Feasible for both mouse and rat models
- ▶ Relatively fast production of founder (F0) animals
- ▶ Dynamic range in transgene expression (e.g. F0s with varied expression patterns or levels)

### Suitability

- ▶ Over(expression) of plasmid- and BAC-based transgenes

### Major Disadvantages

- ▶ Integration(s) can cause deleterious mutations
- ▶ BACs: Passenger genes may provide complication
- ▶ Each integration will vary in location, copy number, transgene structure, and expression profile
- ▶ Possibility for multiple integrations



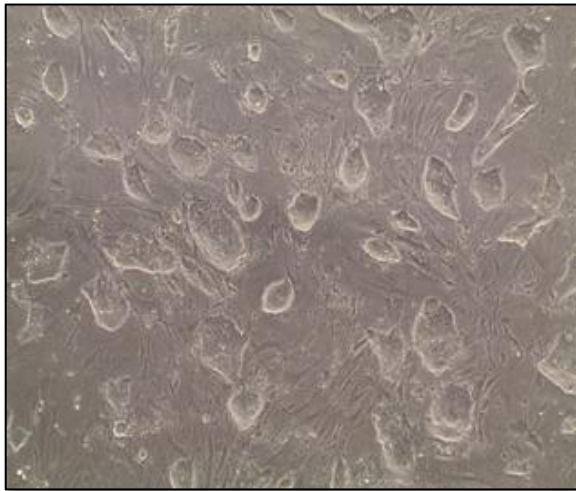
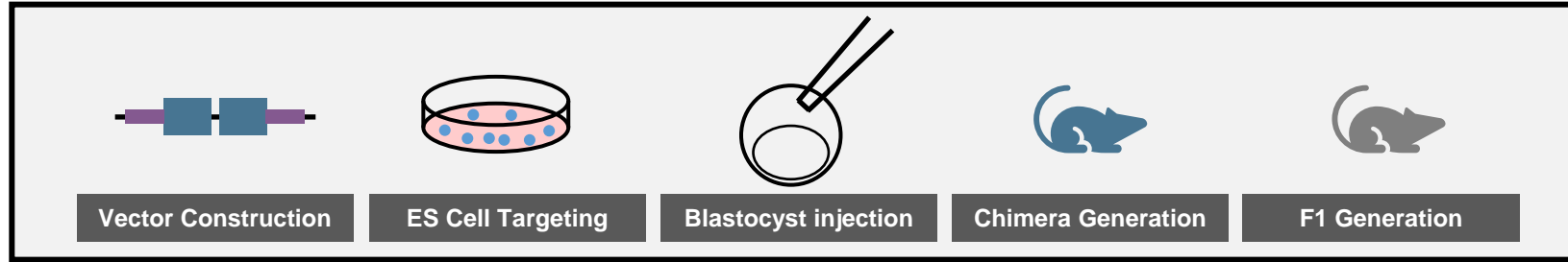
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# Gene Targeting in Embryonic Stem Cells (ESCs)

## Model Generation Process



Mouse ES cells



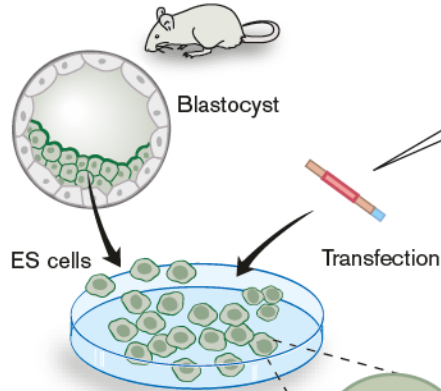
Chimeras

# Gene Targeting in Embryonic Stem Cells (ESCs)

## Step 1 Gene targeting in ES cells

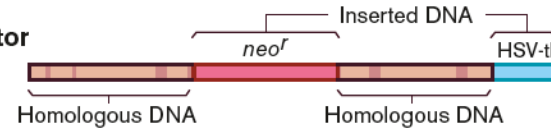
### 1. ES cell culture

Embryonic stem (ES) cells are cultivated from mouse pre-implantation embryos (blastocysts).



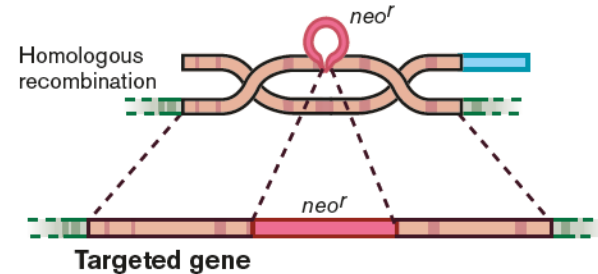
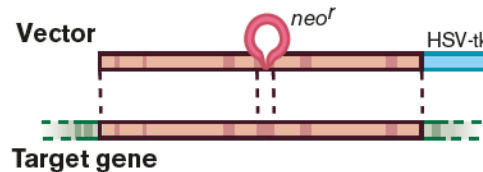
### 2. Construction of targeting vector

The vector contains pieces of DNA that are homologous to the target gene, as well as inserted DNA which changes the target gene and allows for positive-negative selection.



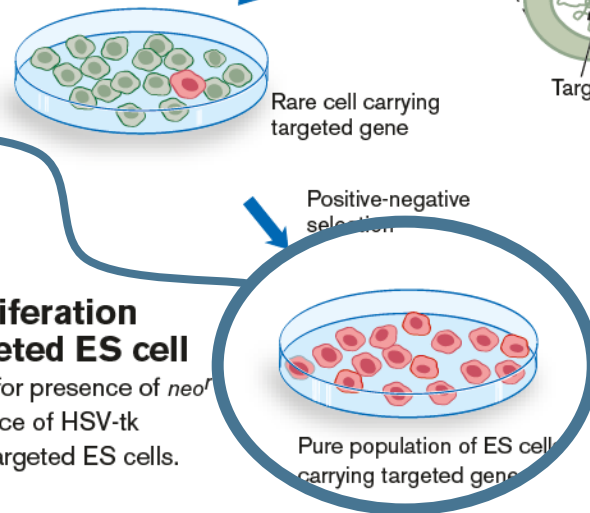
### 3. ES cell transfection

The cellular machinery for homologous recombination allows the targeting vector to find and recombine with the target gene.



### 4. Proliferation of targeted ES cell

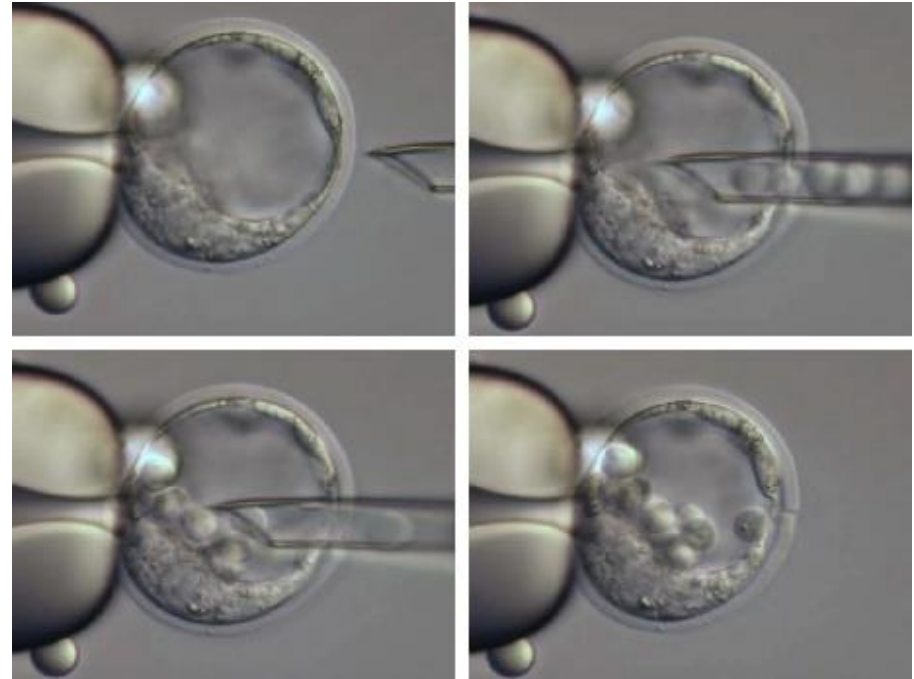
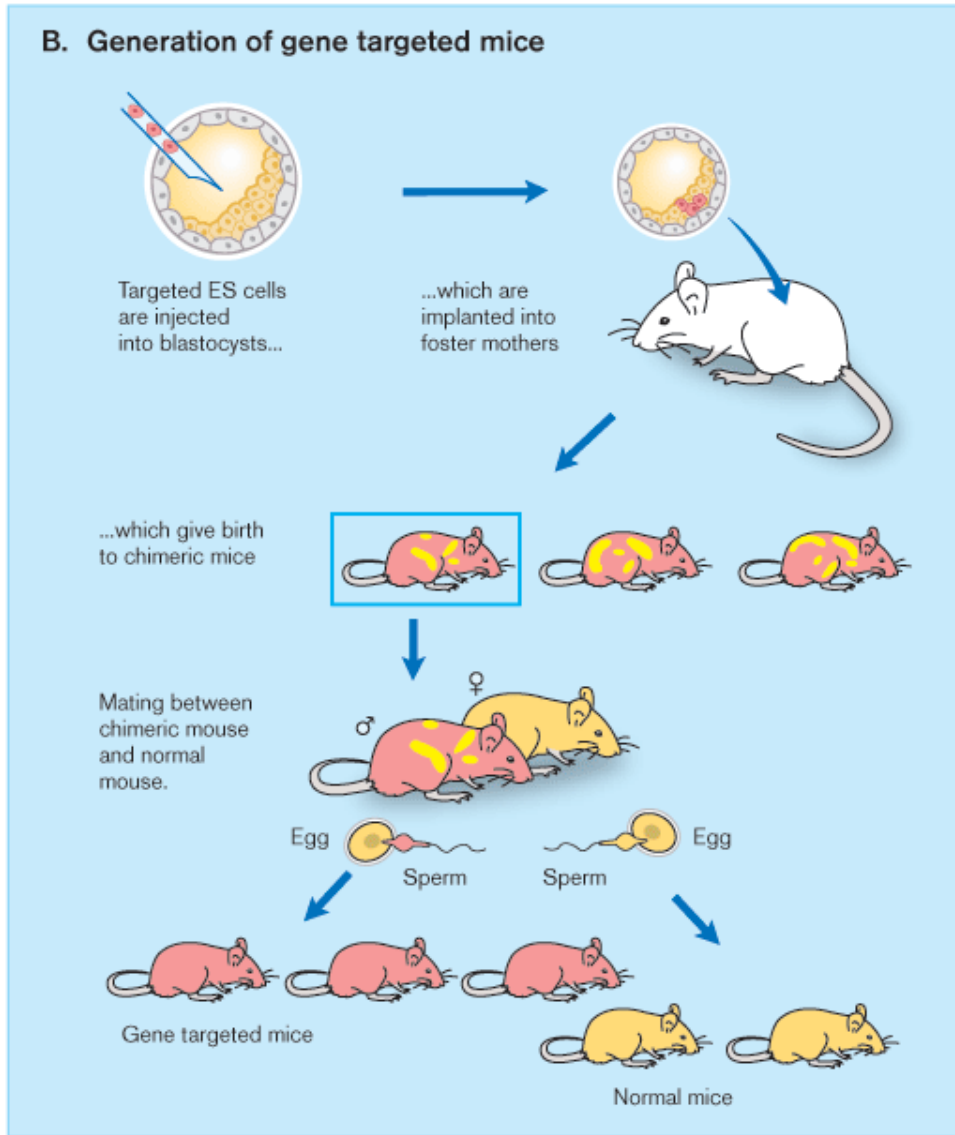
Selection for presence of *neo<sup>r</sup>* and absence of HSV-tk enriches targeted ES cells.



3R

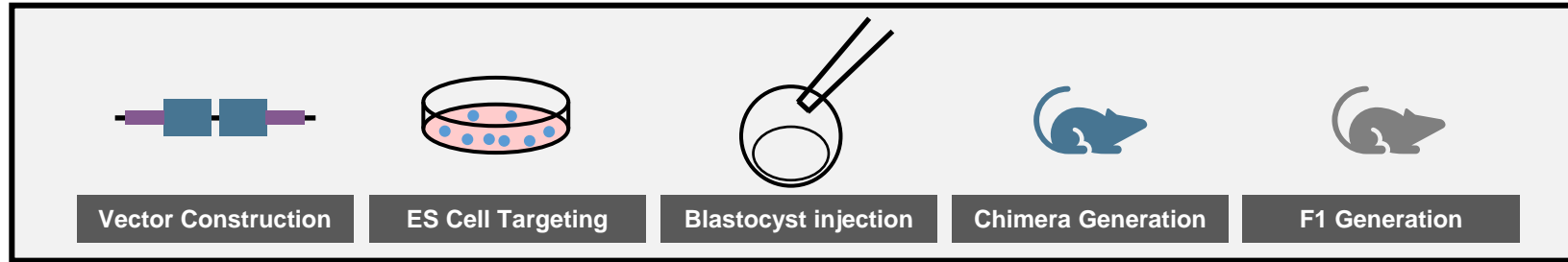
Extensive molecular characterization of multiple clones

# Gene Targeting in Embryonic Stem Cells (ESCs)



# Gene Targeting in Embryonic Stem Cells (ESCs)

## Model Generation Process



### Major Advantages

- ▶ Targeted insertion of a well-defined modification (e.g., single copy, intact structure)
- ▶ Very large and complex modifications possible
- ▶ Extensive and thorough validation (e.g., Southern blots with internal and external probes) prior to generating mice

### Major Disadvantages

- ▶ Limited to mouse model generation
- ▶ Requires an appropriate ES cell line

### Suitability

- ▶ Large and complex modifications (e.g., targeted humanizations >150 kb)

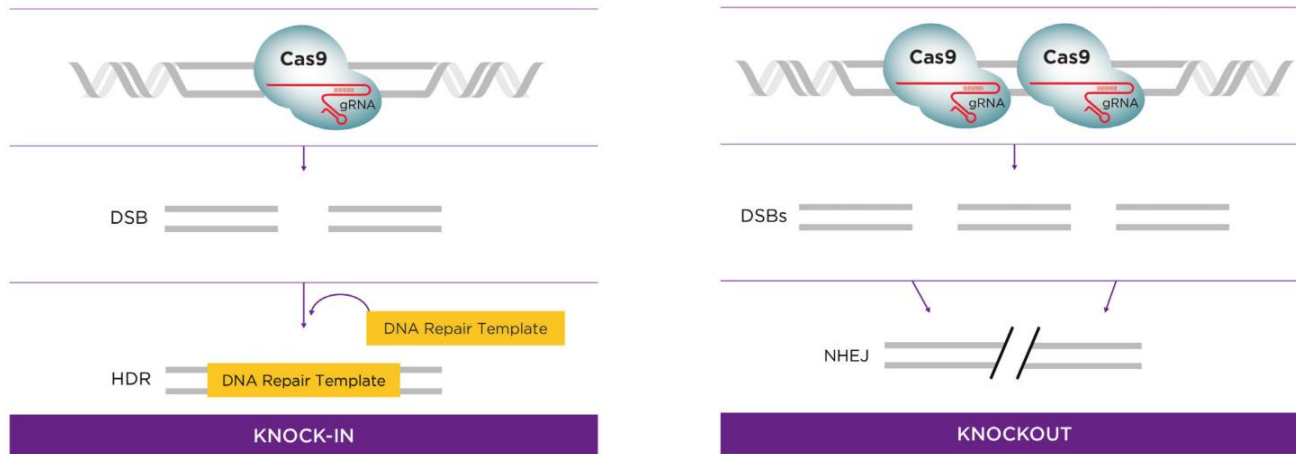
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# Gene Editing in Zygotes Using CRISPR/Cas9

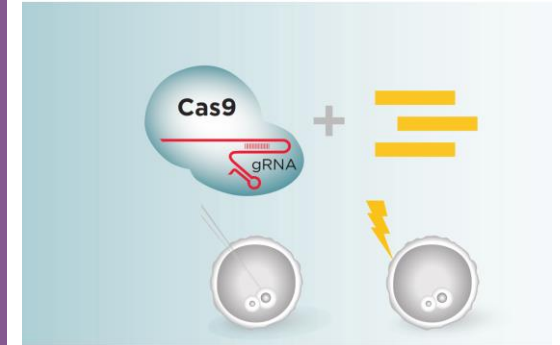
## Model Generation Process



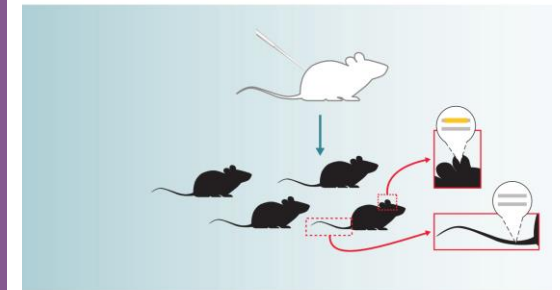
2 wks



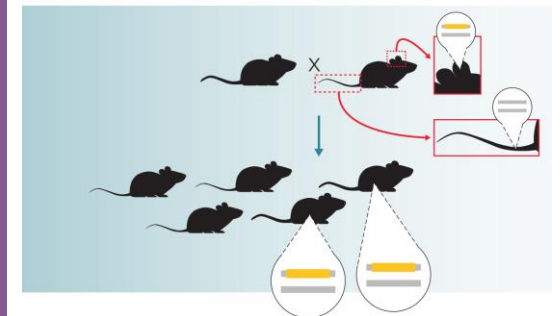
1-2 days



12 wks



12 wks



# Gene Editing in Zygotes Using CRISPR/Cas9

## Model Generation Process



### Major Advantages

- ▶ Feasible for both mouse and rat models
- ▶ Usually fastest timeline to F1 animals
- ▶ Flexibility (e.g., choice of genetic background), ease of design and reagent preparation

### Major Disadvantages

- ▶ Risk for off-target mutations
- ▶ Risk for secondary on-target mutations
- ▶ Different quality control or genetic validation procedures *in vivo* (vs. gene targeting in ES cells)
- ▶ Size of modifications is limited

### Suitability

- ▶ Small and less-complex modifications (e.g., KOs, knock-in point mutations, fluorescent tags, knock-ins <10 kb)



# Choosing the Right Tool for every Model

Targeting efficiency of CRISPR *in vivo* projects vs. ESC-based targeting

	ES Cells	CRISPR
Constitutive Knockout	●●	●●●●
Point Mutation Knock-in	●●	●●●●
Short Tag Knock-in	●●	●●●
Cre Recombinase Knock-in	●●●	●●●
Reporter Knock-in	●●●	●●●
cDNA Knock-in	●●●	●●●
Conditional Knockout	●●●●	●●
Genomic Replacement	●●●●	●

●●●● Excellent ●●● Good ●● OK ● Poor



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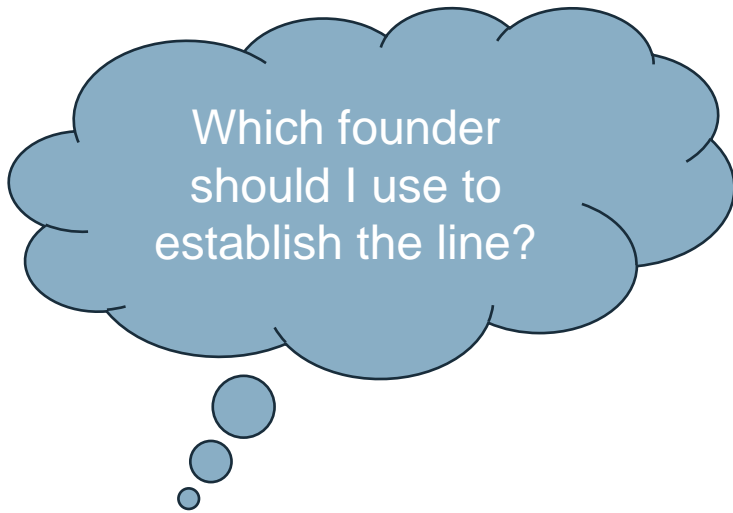
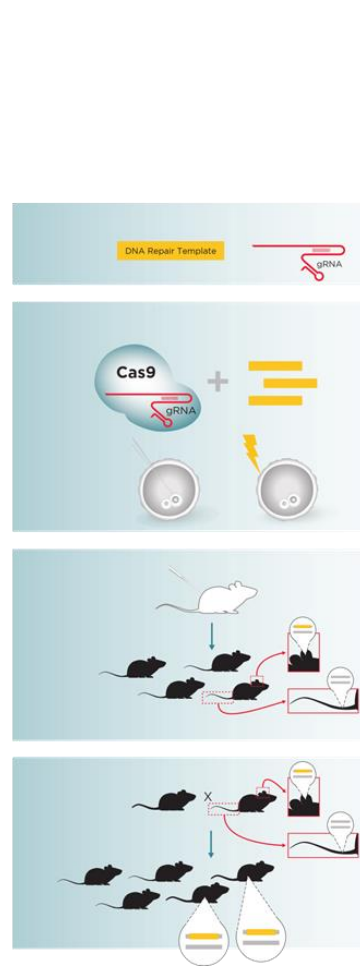
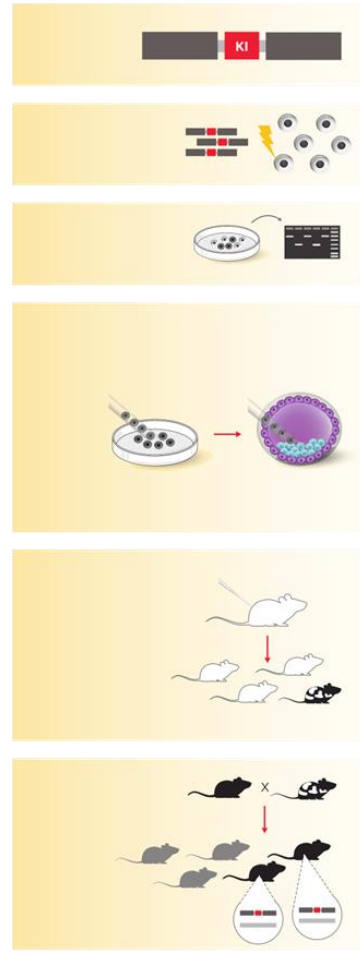
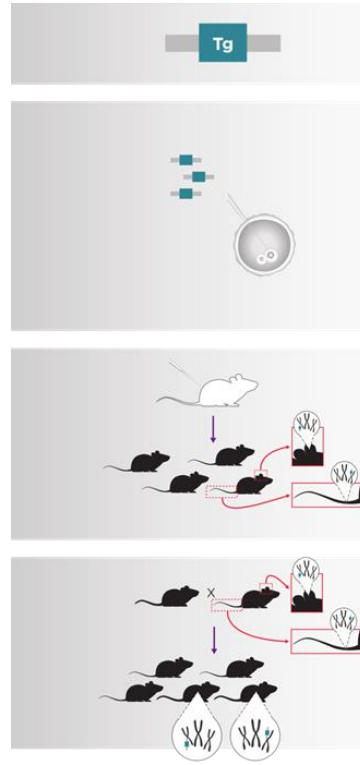
# Advancing Model Generation

The founder challenge

Random Integration Transgenic (RITg)

Gene Targeting in Embryonic Stem Cells

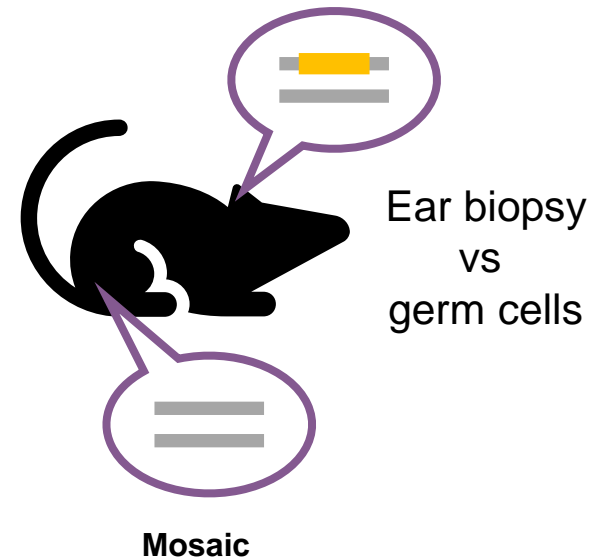
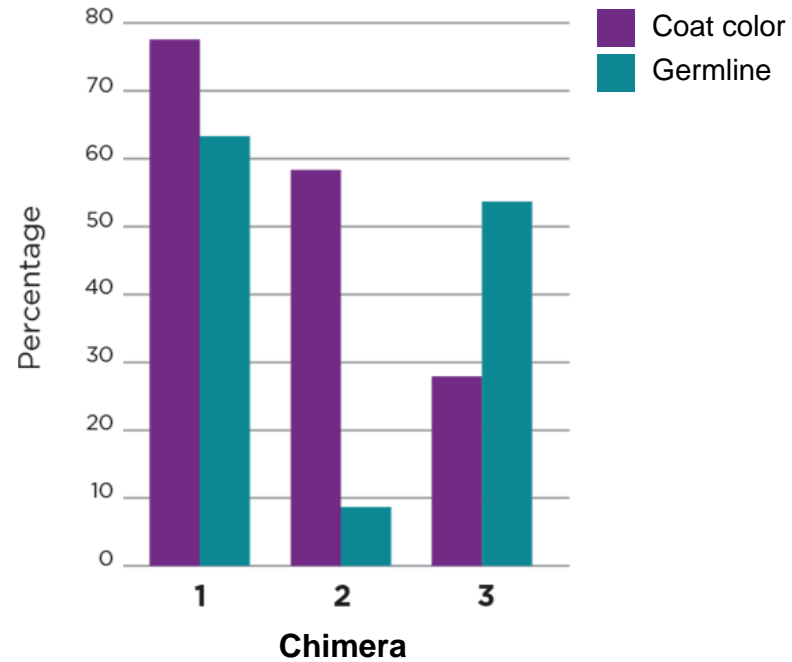
Gene Editing using CRISPR/Cas9



# Advancing Model Generation

## The founder challenge

- ▶ Founders are most likely mosaic or chimeric
  - ▶ Allele frequency in germline unknown



# Advancing Model Generation

## The founder challenge

- ▶ Founders are most likely mosaic or chimeric
  - ▶ Allele frequency in germline unknown
  - ▶ Fertility of founder potentially compromised
  - ▶ RITg: need to breed and analyze multiple sublines



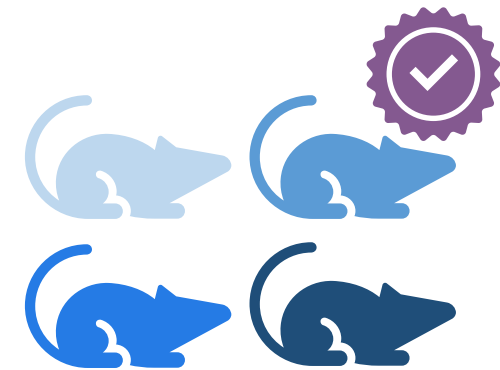
# Advancing Model Generation

Reducing timelines, risks, and surplus animals without compromising quality

- Concept: Data-informed founder selection



	Allele frequency	Expression analysis	Fertility
RITg	Mosaicism (qPCR)	Tissue-specific expression levels (qRT-PCR)	<i>In vitro</i> fertilization
ES Cells	Chimerism (qPCR)	-	<i>In vitro</i> fertilization
CRISPR	Mosaicism (NGS) OT analysis (NGS)	-	<i>In vitro</i> fertilization

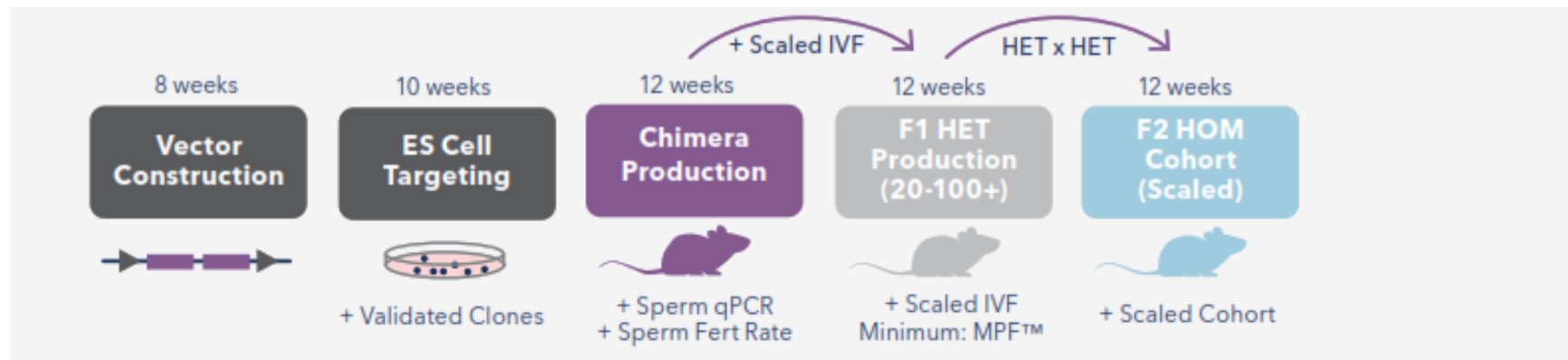
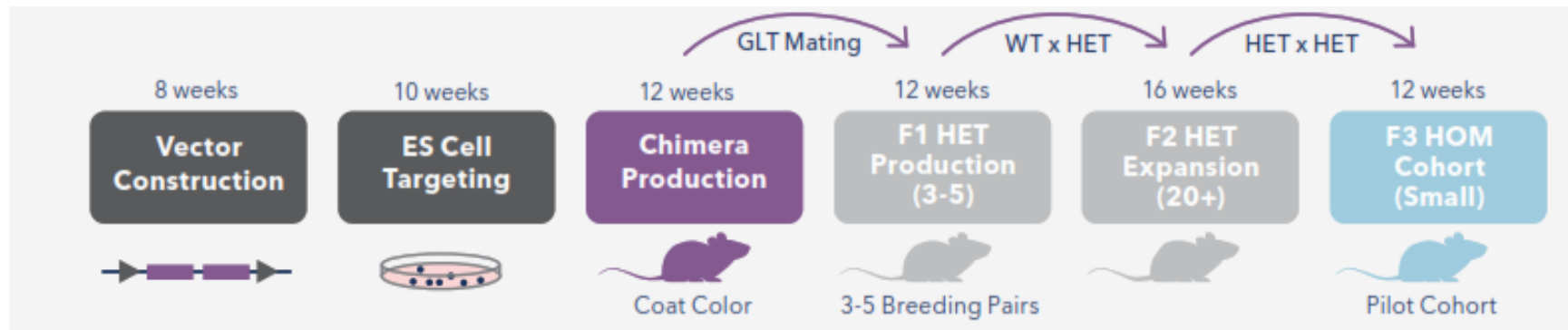


Identify the best founder

# Advancing Model Generation

Reducing timelines, risks, and surplus animals without compromising quality

- Concept: Data-informed founder selection & Faster and scalable generation of F1 animals



# Custom Model Generation

## Summary and Closing Remarks



- The **project goal** determines the strategy and the methodology.
- There are different methods with **their own strengths and weaknesses**.
- There is **not THE ULTIMATE** method for custom model generation.
- **Careful evaluation** of the optimal strategy, methodology and project design is not only important for the research goals, but also for the 3Rs.



**Thank You**



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